

Validation of inertial measurement units to detect and predict horse behaviour while stabled

Katrina Anderson¹  | Ashleigh V. Morrice-West¹  | Elizabeth A. Walmsley¹  |
 Andrew D. Fisher²  | R. Chris Whitton¹  | Peta L. Hitchens¹ 

¹Equine Lameness and Imaging Centre, Melbourne Veterinary School, University of Melbourne, Werribee, Victoria, Australia

²Animal Welfare Science Centre, Faculty of Veterinary and Agricultural Sciences, University of Melbourne, Parkville, Victoria, Australia

Correspondence

Peta L. Hitchens, Equine Lameness and Imaging Centre, Melbourne Veterinary School, University of Melbourne, Werribee, VIC 3030, Australia.
 Email: peta.hitchens@unimelb.edu.au

Funding information

Racing Victoria; University of Melbourne; Victorian Racing Industry Fund of the Victoria State Government

Abstract

Background: Musculoskeletal injuries are observed in Thoroughbred racehorses and may become catastrophic. Currently, there are limited methods for early detection of such injuries. Most injuries develop gradually due to accumulated damage, providing the opportunity for early detection. Horses experiencing pain or lameness may exhibit changes in behaviour so the development of an objective, real-time system monitoring horse behaviour may enable detection of bone injuries before catastrophic failure.

Objectives: To determine whether intensive observational methods of assessing horse behaviour can be replaced by use of inertial measurement units (IMUs).

Study design: Validation study assessing IMU use against video observation.

Methods: Six hospitalised Thoroughbreds (algorithm training data) and 19 Thoroughbred racehorses in-training (algorithm testing data) were equipped with an IMU placed on the lateral side of both forelimbs (left fore, LF; right fore, RF) and monitored in a stable for 4 h. An algorithm was developed to classify behaviour and then validated against video recordings.

Results: Standing was the most prevalent behaviour (LF 88.8%, 95% confidence interval [CI] 88.7–89.0; RF 88.5%, 95% CI 88.4–88.7). IMU classification of recumbent and standing activities showed excellent agreement (sensitivity) with video observation (>98%). This was followed by stepping (LF 89.4%, RF 85.5%) then weight-shifting (LF 54.3%, RF 61.5%). Predictions from the algorithm showed misclassification of 2.5% (LF 5500/225 352, RF 5218/210 170). Excluding standing, misclassification was 6.8% (1705/25 158) and 7.5% (1812/24 077) for the left and right forelimbs, respectively, with pawing and weight-shifting most frequently misclassified.

Main limitations: Increasing the number of horses and types of behaviours observed may improve predictions.

Conclusions: IMUs displayed a high sensitivity to movement on a small number of horses, and with further validation they have the potential to effectively monitor behaviour of racehorses in training. However, more sensitive methods may be

This is an open access article under the terms of the [Creative Commons Attribution-NonCommercial](https://creativecommons.org/licenses/by-nc/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes.

© 2022 The Authors. *Equine Veterinary Journal* published by John Wiley & Sons Ltd on behalf of EVJ Ltd.

needed to validate the classification of weight-shifting behaviour. Future studies should evaluate the association between each behaviour and musculoskeletal injury.

KEYWORDS

horse, IMU, orthopaedic pain, racehorse, Thoroughbred, video monitoring

1 | INTRODUCTION

Musculoskeletal injuries are the most common cause of lost training days and early retirement in racehorses, resulting in substantial economic impact to owners and trainers.^{1,2} Most injuries in racehorses are due to tissue fatigue, developing gradually as bone or tendon integrity is overcome by repetitive load.³ For bone, this is evidenced by post-mortem examinations revealing pre-existing pathology for fracture including periosteal callus formation, focal bone resorption and microdamage.^{4–6} Because most injuries develop over time, there is an opportunity for early detection.^{4,6} The use of diagnostic imaging techniques to detect early injury has been investigated, but low specificity, high cost and logistics limit their use for screening large numbers of horses.^{7–10} Additionally, a trend towards increased racing stable size (number of horses per trainer) over the past decade makes close monitoring of horses more challenging.^{11,12} Continuous observation is not feasible nor practical for most racing stables, and horses tend to hide their discomfort in the presence of observers.¹³ Therefore, an automated, objective monitoring system to aid in the assessment of racehorse behaviour is needed.

Orthopaedic pain, commonly expressed as lameness in horses, is also associated with changes in behaviour including weight-shifting,¹⁴ pawing¹⁵ and restlessness.¹⁶ The development of an objective, preferably real-time, system to monitor horse behaviour may enable the early detection of bone injuries before more overt clinical signs developing. Numerous measures to evaluate pain have been developed to score severity based on ordinal or simple descriptive scales, including composite multifactorial scales and facial expressions.^{15–24} Horses in pain are shown to spend more time inactive and those with known orthopaedic pain have been observed unloading one limb, pawing and weight-shifting, although behaviours such as pawing are not specific to orthopaedic discomfort.^{14–17} Further, rather than singular expressions of behaviour in horses, composite and facial expression-based pain scales show promise, at least for acute pain.²⁵ A combination of behaviours (posture, weight-bearing) and physiological signs (temperature, heart rate) were found to be associated with movement asymmetry from the Equine Pain Scale and Composite Pain Scale.²⁶ Subtle signs of orthopaedic pain have also been recognised in a first attempt using pose estimation of raw video data, trained on a data set of facial and upper body poses in horses with acute experimental pain.²⁷ However, current methods of intermittent monitoring may not accurately detect the frequency of pain-related behaviours.¹⁷

Wearable inertial measurement units (IMUs) use orientation and acceleration to interpret activity.^{28–31} IMUs have been used to detect Parkinson's disease in humans,³² and to study movement of marine,

terrestrial and airborne animals.^{33–36} Movement is recorded as acceleration signals and analysed by machine learning tools to identify patterns that can be used to discriminate behaviours. Algorithms have been developed and validated to classify dog behaviour with excellent agreement (>0.90),³⁷ detect lameness in sheep with an 82% accuracy,³⁸ monitor lying behaviour in cattle³⁹ and predict foaling in broodmares.⁴⁰ More recently, IMUs have been used to predict step count in horses under stall confinement with excellent agreement (>0.99),⁴¹ and assess changes in postural sway for horses with induced lameness.⁴²

Effective remote monitoring of horses could be used in racing stables to detect subtle changes in behaviour. The objective of this study was to determine whether IMUs are sensitive enough to detect horse behaviours when compared with intensive methods, such as constant observation, in what would be considered a healthy population, as orthopaedic pain can be sub-clinical. We aimed to develop an algorithm to classify horse behaviour and (1) determine the accuracy and precision of the behavioural events predicted by IMUs; and (2) validate their use to quantify posture and behaviour, specifically, in detecting static orientations of standing and recumbency as well as dynamic movements of stepping, pawing and weight-shifting.

2 | MATERIALS AND METHODS

2.1 | Study population and environment

Twenty-five Thoroughbred horses were recruited from the University of Melbourne Equine Centre (UVet) ($n = 6$) and 1 Victorian registered racehorse trainer ($n = 19$). Six horses recruited from UVet were utilised for algorithm development (mean age 3.7 years, standard deviation (sd) 2.2 years). We reasoned that this population would allow for observation of horses likely to be exhibiting signs of pain and/or discomfort in a closely controlled environment. For validation, the developed algorithm was then tested using video recordings of 19 horses in a typical racehorse training stable environment that met the inclusion criteria; (1) Thoroughbred racehorse; (2) ≥ 3 years and (3) deemed fit for race training. This test population consisted of $n = 3$ females, $n = 13$ geldings and $n = 3$ colts and stallions, with a mean age of 3.8 years (sd 1.07).

Horses were confined to individual 360×370 mm² stables for the recording periods. Stables had visual access to neighbouring horses by means of open metal bars dividing the sides of stalls (above ~ 1.3 m of solid wall) and on the front wall inwards to a breezeway. Rear walls were solid with no visible distractors. Ad-lib hay and water were available and positioned either on the ground or at chest height.

Horses were provided with hard feed twice daily, morning and late afternoon.

2.2 | Sample size analysis

Estimation of sample size was based on a previously published validation of motion sensors in dairy cattle that identified lying, standing and moving (four calves in training data set; five calves in validation data set). As that study found that the motion sensor accurately measured high-prevalence behaviours, but less accurately measured low-prevalence behaviours, we increased the number of horses used (6 training; 19 validation) for this current study.⁴³

2.3 | Accelerometer data

Horses were equipped with two 500 Hz nine axis IMUs (length: 40 mm; width: 28 mm; depth: 15 mm; mass: 12 g; accelerometer range: ± 16 g; gyroscope range: ± 2000 degree per second ($^{\circ}\text{s}^{-1}$); Vicon). Sensors were positioned on the lateral side of both metacarpi (left fore, LF; right fore, RF), secured in a horse boot with a purpose-built Velcro pouch (Figure 1).

The duration of data collection was limited to the battery life of the sensors (<5 h). Therefore, data were collected in varied 4 h time windows, ranging from start times of approximately 10:00–22:00, as dictated by stable activities and horse availability, avoiding active periods such as trackwork, with 8/19 horses being fed within the time window that they were monitored. A mobile application (IMeasureU, Vicon) was used to start and stop data capture.

2.4 | Video analysis

Video footage was recorded using a video camera (TECHview 1080p) mounted to a bracket that was securely fastened to the stable wall in one corner at a height of 2.4 m, which allowed for a view of the entire stable (101°). The mean footage for each horse was 224 min (sd 37.6). The sensors were tapped by hand multiple times in view of the camera to provide a time stamp that was used to synchronise the video footage with the IMU data.³⁰

2.5 | Data processing and algorithm development

Data processing and algorithm development was conducted using MATLAB version R2018a (MathWorks). MathWorks, Natick, Massachusetts, United States.

Sensor data were extracted from the IMUs and imported into MATLAB. A custom-designed MATLAB script was used to process the data. The z- and x-axes from the sensor of the left forelimb were multiplied by -1 to convert data equivalent to that of the right forelimb.

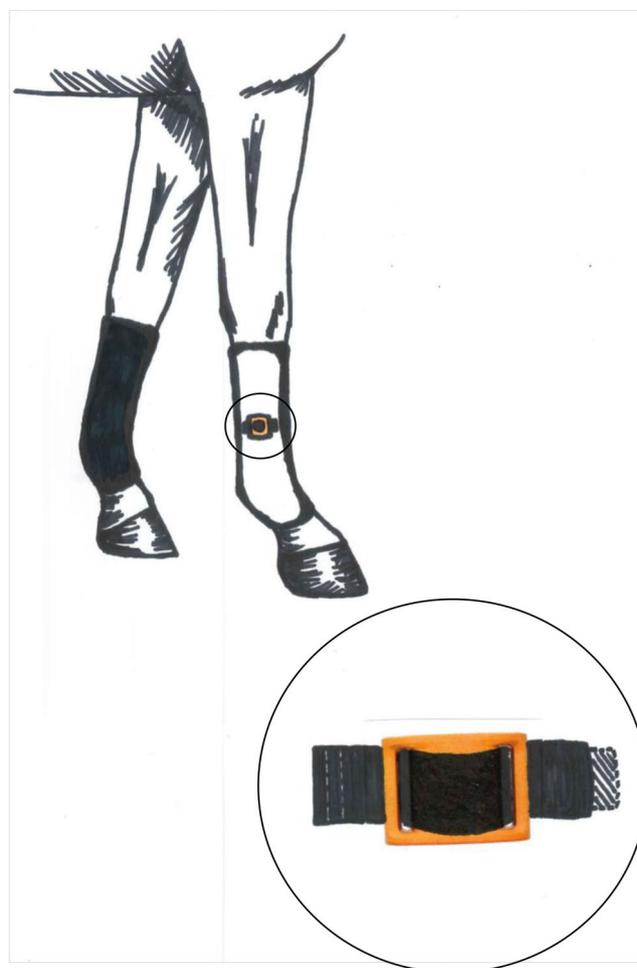


FIGURE 1 Depiction of boots with inertial measurement unit (IMU) placement. Sensors were positioned on the lateral side of both metacarpi (left and right fore), secured in a horse boot and stitched into a purpose-built Velcro pouch (inset).

Data were collected from hospital-admitted horses ($n = 6$) and used to train the algorithm (training data). The video recording was used to manually label time frames from sensor data to a behaviour class. The time stamp allowed one observer (KA), with experience working in Thoroughbred racing stables and as a post-graduate trained in equine behavioural annotation, to coordinate IMU time with video footage. For each movement on IMU, the start time was recorded and labelled according to the behaviour observed on video in 1-s intervals.²⁹ This time interval was chosen because it has previously been reported to be sensitive to detection of movement for most behaviour classes.³⁴

We defined six basic behavioural classes. Each behaviour was encoded from 1 to 6: (1) left recumbency; (2) stepping; (3) pawing; (4) standing; (5) weight-shifting; and (6) right recumbency (Table 1). Behaviours were recorded based on the 1-s time-sampling interval. For recumbency, stepping, and standing they were based on previous studies comparing IMU recordings to video observations in cattle. Recumbency was defined as the animal either in lateral or sternal recumbency for the entire 1 s; and stepping as a minimum of one

complete forward or backward progressive step.^{44,45} Pawing was defined as pawing the ground or air or pointing or hanging the limb, and weight-shifting as changes in weight distribution or shifts, muscle tremors or non-weight bearing of either limb per Bussi eres et al.¹⁵

Each classified behavioural observation was allocated to a row number which was found by multiplying the time in seconds by 500 (because the data was recorded at 500 Hz; each second contained 500 points). Five features were calculated using a moving

window calculation for the time interval across the entire array; including mean, median, minimum, maximum and sd. The five features were calculated for each of the eight columns, consisting of six acceleration signals (acceleration x, y, z; gyroscope x, y, z) and an additional two resultant vectors calculated using the Pythagorean method. These calculations were allocated to a behaviour and used to train machine learning classifiers. Each classifier was evaluated using 10-fold cross-validation and the accuracy of each was determined.⁴⁶

TABLE 1 Description of broadly classified behaviours observed on video recordings in $n = 25$ Thoroughbred racehorses stabled at rest

Behaviour	Description
Left recumbency	The horse is in sternal or lateral recumbency with its left side down for >3 s, including behaviour such as rolling.
Stepping	Progressive movement of hoof from one location to another one.
Pawing	Non progressive movement. Continuous action of digging with one limb with the horse otherwise stationary.
Standing	Standing on both forelimbs without unloading or moving (≥ 1 s)
Weight-shifting	Non progressive movement. Action of unloading weight from one limb to another (not necessarily taking a limb off the ground), often continuous between left forelimb and right forelimb.
Right recumbency	The horse is in sternal or lateral recumbency with its right side down for >3 s, including behaviour such as rolling.

TABLE 2 Summary information about the observed behaviour sequences of both left and right forelimb of $n = 19$ individual Thoroughbred horses in race training monitored in a stable

	Left limb observed		Right limb observed	
	Mean (sd)	Min, max	Mean (sd)	Min, max
Total length of observation (s)	12 093 (2149)	8512, 16 214	11 886 (2095)	8415, 14 763
No. of transitions	448 (165)	231, 982	431 (140)	230, 725
States observed (max = 6)	4 (1)	3, 6	4 (1)	3, 6
Duration of each behaviour (%) ^a				
Left recumbency (8/19)	4.4 (8.2)	0, 29.8	4.7 (8.5)	0, 29.9
Stepping	3.5 (2.0)	1.5, 9.8	3.5 (2.0)	1.8, 9.4
Pawing	0.1 (0.1)	0, 0.2	0.2 (0.2)	0, 0.7
Standing	89.0 (7.7)	66.6, 97.9	88.7 (7.6)	66.7, 97.3
Weight-shifting	1.2 (0.7)	0.1, 2.6	1.3 (0.8)	0.1, 3.5
Right recumbency (4/19)	1.9 (3.9)	0, 11.2	1.8 (4.0)	0, 12.4
Entropy ^b	0.22 (0.1)	0.06, 0.42	0.23 (0.1)	0.08, 0.42
Complexity ^c	0.09 (0.1)	0.04, 0.16	0.009 (0.02)	0.05, 0.13

^aThe percentage of time spent in each state of behaviour during the observational hours, which include weight shifts before and after stepping which were excluded from analysis.

^bA measure of diversity of states that compose the sequence, based on time spent at different states (where 0 = one behaviour observed and 1 = equal time spent at each state).

^cCombines entropy and number of transitions.

2.6 | Validation

The developed algorithm was used to predict behaviour in $n = 19$ horses in race training (unlabelled test data set) using patterns recognised from the previously labelled training data set.^{36,46} The predicted behaviours from the algorithm were blinded to the observer (KA), who manually classified behaviour per second from the video to one of the six defined behaviours. If behaviour could not be recorded because the limb of interest was obscured, the data was recorded as missing. Abnormal behaviour, that is, if activity could not be classified (e.g., sudden movement to the right), was recorded and excluded from analysis. Observations where the sensor slipped, malfunctioned, or did not record data (all observations for the right forelimb of one horse [HorseID 18] and $n = 313$ observations across various horses) were also excluded from analysis.

An internal-clock drift in the IMU was corrected using a time stamp to detect the difference between true and IMU time. Each sensor was manually calibrated at consistent intervals based on the correction factor determined. Since the time resolution of the device was 1-s, the correction was implemented in the data as

occasional repeats of the same time, or occasional 1-s skips, as previously reported.⁴⁷

The transitional period between standing and stepping was frequently classified as weight shifts (LF 55.8%, 4087/7316; RF 50.7%, 3623/7143 of total weight shifts). To account for this misclassification, a variable to exclude weight shift movements predicted before and after stepping behaviour was generated. A period of 5 s following the transition of recumbent to standing was not used in validation analysis because behaviour displayed could not be defined.

2.7 | Data analysis

Statistical analyses were conducted in Stata/IC version 15.0 (Stata-Corp).⁴⁸ The unbalanced ratio of behaviours meant the accuracy for all classifications was excellent and not informative to the overall performance of the algorithm. Therefore, we evaluated the performance of the algorithm based on its ability to correctly predict individual behaviours, including sensitivity and precision, as previously done by Martiskainen et al.⁴⁹ A confusion matrix describing the performance of the algorithm at classifying behaviours was generated.⁴⁶

Sensitivity (true positive/[true positive + false negative]), and precision (true positive/[true positive + false positive]) were determined to assess the accuracy of the algorithm to predict horse behaviour in the test data ($n = 19$), by comparing the video observation to the behaviour predicted. Continuous data were assessed using the Shapiro–Wilk test, and as the majority were normally distributed, all data were reported as mean and standard deviation. Sequence index plots were generated in Stata (StataCorp). Sequence analysis was conducted in R using the package TraMineR (R Foundation for Statistical Computing).^{50,51} The duration of each behaviour was calculated as a mean of observed behaviour for each behaviour. For recumbency behaviour, the duration percentage was a mean calculated from only those horses that were recumbent at least once.

3 | RESULTS

3.1 | Training data

The performance of the behaviour classifier algorithm on the training data is presented as a confusion matrix in Table S1, with sensitivity ranging from 80.3% to 96.7% and precision from 85.9% to 100% for each behaviour. Left recumbency had the highest precision, followed by standing, pawing, weight-shifting, stepping and right recumbency. Stepping and pawing had the lowest sensitivity.

3.2 | Test data

Tables 2 and S2 describe the summary data recorded from 19 individual horses for observations, transitions and duration of behaviours.

TABLE 3 Total agreement between predicted and observed behaviour $n = 19$ Thoroughbred stabled horses over a 4-h monitoring session per second. Predicted behaviours were classified using inertial measurement units placed on the left fore (LF) and right fore (RF) metacarpi of each horse. Observed behaviours were classified using video recordings.

Predicted	Observed													
	Left recumbency		Stepping		Pawing		Standing		Weight-shifting		Right recumbency		Total	
	LF	RF	LF	RF	LF	RF	LF	RF	LF	RF	LF	RF	LF	RF
Left recumbency	11 477	11 466	1	0	0	0	0	0	2	3	1	3	11 481	11 472
Stepping	13	13	5805	5075	31	56	1113	895	477	377	1	0	7440	6416
Pawing	2	1	29	9	79	56	13	3	4	6	1	0	128	75
Standing	2	1	441	528	3	0	196 399	182 687	479	483	0	0	197 324	183 699
Weight-shifting	0	8	216	323	1	1	2669	2508	1143	1391	1	0	4030	4231
Right recumbency	0	0	0	0	0	0	0	0	0	0	4949	4277	4949	4277
Total	11 494	11 489	6492	5935	114	113	200 194	186 093	2105	2260	4953	4280	225 352	210 170

Behaviours not classified within the six classes, behaviour not observed, and behaviour detected in the second before and second after 'stepping', were excluded from calculations of agreement. Other behaviours not analysed included: shake, roll, kick, lift leg and scratch leg ($n = 935$ s).

Video observation detected standing as the most dominant behaviour pattern in the test data set, with prevalence of 88.8% and 88.5% for the left fore and right fore, respectively (Tables 3 and 4), followed by recumbency (left-sided recumbency prevalence of 5.1% and 5.5% and right-sided 2.2% and 2.0%) and stepping (2.9% and 2.8%, respectively) (Table 4). Over the complete observation period, 57.9% of horses (11/19) were recumbent for a period of more than 3 s, most of which occurred during the afternoon (between 3.00 PM and 5.00 PM) or evening (after 8.00 PM) (Figure 2). The mean total recumbent time for all horses was 14.5 min (sd 19.4; range 0–70.9). One horse was recumbent for 27.5% of the observational period (70.9/257.6 min).

Greater than 60% of behaviour states observed were followed by the same behaviour state in the next recorded second, with the exception of weight-shifting (Table S3). Weight-shifting was most often followed by standing (47%) or more weight-shifting (42%). Calculations of transition rates when repeated events in sequence were ignored showed a high probability that stepping (90%) and weight

shifts (81%) were followed by standing. The longitudinal characteristics of sequences between horses are illustrated (Figure 2), which can be used to interpret how individual horses change behaviour state over time.

The total agreement between predicted and observed behaviours (Table 3) was used to calculate precision (proportion of positive predictions that were correct) and sensitivity (proportion of the positive cases that were predicted positive) of the algorithm to predict each behaviour classification, stratified by limb (Table 4). Overall, precision varied from 28.4% (weight-shifting) to >99.9% (recumbency), and sensitivity varied from 49.6% (pawing) to 99.9% (recumbency). Standing and recumbency were predicted with a precision and sensitivity of >98% in both left and right limbs. Stepping behaviours were predicted with 78.0% and 79.1% precision and 89.4% and 85.5% sensitivity for the left and right forelimb, respectively. Compared with video observations, after exclusion of 1 s before and after stepping and abnormal observations, the predictions from the algorithm showed mean misclassification by horse of 2.6% for the left and right forelimb (LF sd. 1.6%, range 1.0, 7.8%; RF sd. 1.4%, range 1.2, 6.7%). Overall misclassification was 2.4% (5500/225 352) and 2.5% (5218/210 170) for the left and right forelimbs, respectively (total 2.5%). However, excluding standing (the most prevalent behaviour), overall misclassification was 6.8% (1705/25 158) and 7.5% (1812/24 077), respectively. Pawing

Behaviour	Sensitivity (95% CI)			
	Left forelimb	95% CI	Right forelimb	95% CI
Left recumbency	99.9	99.8–99.9	99.8	99.7–99.9
Stepping	89.4	88.6–90.2	85.5	84.6–86.4
Pawing	69.3	60.0–77.6	49.6	40.0–59.1
Standing	98.1	98.0–98.2	98.2	98.1–98.2
Weight-shifting	54.3	52.1–56.4	61.5	59.5–63.6
Right recumbency	99.9	99.8–100.0	99.9	99.8–100.0
Behaviour	Precision (95% CI)			
	Left forelimb	95% CI	Right forelimb	95% CI
Left recumbency	>99.9	99.9–100.0	99.9	99.9–100.0
Stepping	78.0	77.1–79.0	79.1	78.1–80.1
Pawing	61.7	52.7–70.2	74.7	63.3–84.0
Standing	99.5	99.5–99.6	99.4	99.4–99.5
Weight-shifting	28.4	27.0–29.8	32.9	31.5–34.3
Right recumbency	>99.9	99.9–100.0	>99.9	99.9–100.0
Behaviour	Prevalence (95% CI)			
	Left forelimb	95% CI	Right forelimb	95% CI
Left recumbency	5.1	5.0–5.2	5.5	5.4–5.6
Stepping	2.9	2.8–3.0	2.8	2.8–2.9
Pawing	0.1	0.0–0.1	0.1	0.0–0.1
Standing	88.8	88.7–89.0	88.5	88.4–88.7
Weight-shifting	0.9	0.9–1.0	1.1	1.0–1.1
Right recumbency	2.2	2.1–2.3	2.0	2.0–2.1

TABLE 4 Sensitivity, precision and prevalence as percentages and their respective 95% confidence intervals (95% CIs) of behaviours between video observation and algorithm prediction of behaviour in $n = 19$ stabled Thoroughbreds, stratified by limb

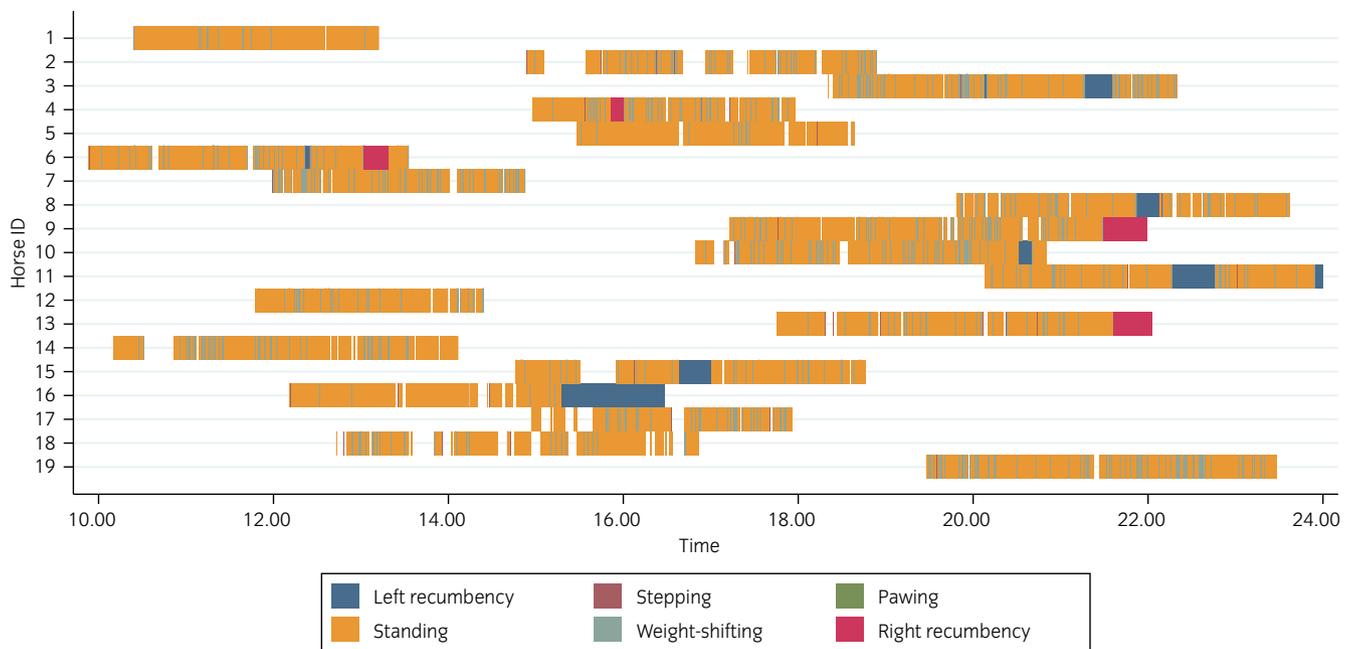


FIGURE 2 Sequence index plot demonstrating changes in behaviour states over time of Thoroughbred racehorses in training ($n = 19$). Changes in behaviour state are shown by changes in colour.

was most often misclassified as stepping (LF 27.2% and RF 49.6% of true positives). Weight-shifting was most often misclassified as either standing or stepping (LF 45.4%; RF 38.1% of true positives).

4 | DISCUSSION

Using IMUs, an algorithm was developed and validated to accurately identify horse behaviour, and therefore has the potential to replace current methods of observation and enable continuous monitoring. Using a supervised learning algorithm, we achieved >85% sensitivity for four behaviours: left recumbency, right recumbency, stepping and standing; and 50% to 69% sensitivity for pawing and weight-shifting. The overall performance of the device suggests it could be developed into a system able to monitor Thoroughbred racehorses in their stables, and over time may allow detection of subtle changes in behaviour patterns.

The horses in our study spent the greatest time standing (89%) and the least time pawing (<1%). A previous study observing stabled horses for 72 h (divided into 12-h segments over 6 days, with up to 4 h paddock time per day) found the majority of time was spent standing (55%) and grazing (24%). Assuming horses were standing while grazing, this totals 79%.⁵² Our findings may be similar, though we did not independently categorise eating behaviours. Horses have been observed to spend most of their recumbent time in sternal recumbency rather than on their side.⁵² We were able to differentiate between left and right lateral recumbency but we did not classify sternal recumbency separately.

The sensitivity for detecting recumbent and standing in this study was excellent (>98%), comparable to previous validation studies in

cattle and horses ($\geq 98\%$).^{44,53,54} However, sensitivity for predicting pawing was much lower, with the right forelimb sensor correctly predicting only half of observed pawing episodes. Pawing movements can be brief, so mismatches between the sensor and human observation time recordings are more likely than for behaviours of a longer duration. Similar validation studies found that after slowing the recordings down to review disagreements, the device predicted correctly, and the observer was incorrect.³⁷ In our study, the video footage regularly replayed at 50% of real time, along with the recording of behaviour every second, was intended to help identify these rapid movements. Recently, a small study using observation and convolutional neural networks, found the largest contributors to a lower accuracy were the misclassifications of pawing, rolling and flank watching.⁵⁵ These behaviours were only performed by one horse during the collection of training data and therefore made up only 2.4% of the data set. The infrequent display of pawing limits training opportunities for classifiers. Additionally, individual pawing patterns were diverse, producing distinctive acceleration patterns for each horse, unlike the repetitive movement pattern observed during walking or recumbency. Therefore, larger input data sets are required for training algorithms to recognise pawing behaviour.

We placed the motion sensors on the forelimbs. This position was previously shown to achieve the highest accuracy for discriminating between equine gaits, including walk, trot and canter.⁵⁶ Placement on the right forelimb vertical axis has been demonstrated to have excellent agreement (ICC 1.0) between sensor-based step count and video-based step count for horses confined to stalls.⁴¹ Placement of a motion sensor around a cow's neck has been successful for recognising rumination and eating behaviours,^{49,57-59} but sudden head movements can disrupt recognition of other behaviour patterns and collars

may move freely, independent of animal movement.⁵⁸ Additionally, sensors placed on the neck have difficulty differentiating standing and sternal recumbency because the orientation of the axis does not change.⁴⁹ Forelimb placement, as in the present study, can differentiate these postures using the perpendicular difference between x- and y-axes.⁴⁵ Robert et al. placed the motion sensor on the hindlimb of cattle and reported excellent agreement with video for lying and standing (99.2% and 98%, respectively).⁴⁴ However, walking classification accuracy was significantly lower (67.8%). Sensors placed on the ear, collar and leg yielded different prediction accuracy for lameness in sheep. Leg-mounted sensors cannot differentiate standing from grazing, unlike ear-mounted sensors which can distinguish non-grazing standing with a 96% prediction accuracy.³⁸ As eating and drinking were not classified in this current study, concurrent behaviours, particularly those like eating hay could confound the detection of weight-shifting. Additional sensors strategically placed could be utilised to detect a movement pattern unique to one behaviour that discriminates it from others. Although multiple sensors would not be practical in a racing stable, further investigations would be required to determine if different sensor placement could improve prediction accuracy. Future work could include classifying more behaviours that are performed by horses and associated with stress or discomfort such as eating behaviour or time spent in sternal or lateral recumbency, physiological signs, additional placement of IMUs on hindlimbs to potentially improve detection of weight-shifting behaviour, body behaviours (including posture, head position, location in the stable, focus, and interactive behaviour), or integration of other algorithm-based pain assessment scales such as those used for facial assessment of pain.

The algorithm showed 54% and 62% sensitivity for left and right forelimbs, respectively, in correctly predicting observed weight shifts, however, precision was only 28 and 33%, respectively. Misclassified weight shifts predicted by the algorithm were most commonly recorded by the observer as standing. This may be due to the high sensitivity of the IMU system to detect small accelerations (sampling rate: 500 Hz) compared with the temporal resolution of the human eye (15–20 Hz).⁶⁰ It is difficult to determine true sensitivity when the gold standard is subjective and objective tools have shown higher sensitivity to detect clinical signs before human diagnosis.^{61,62}

Weight-shifting was also more difficult to classify when compared with behaviours that occurred over longer periods of time without change. The high probability of a transition between stepping, standing and weight-shifting (Table S3) may account for the difficulty in differentiating between individual weight shifts as an independent (primary) movement or as a transitional (secondary) behaviour. These three behaviours in particular can be both a primary behaviour or secondary element of another behaviour, and thus in future studies it is important to distinguish between progressive stepping such as walking from one locale to another or nonprogressive stepping that is associated with, for example, weight-shifting or pawing; or pawing prior to rolling or recumbency. In an attempt to label weight-shifting where it was deemed a primary behaviour, we excluded weight shifts

1 s before and 1 s after stepping behaviour, although this time period may not effectively account for all transient detections of weight shifts (such as before and after pawing). Therefore, it may be necessary to make improvements to the algorithm to increase sensitivity for weight-shifting as a primary movement, or to label these behaviours separately in the context of being primary or secondary. Further, behaviours occurring concurrently to those recorded by the IMU, but that were not accounted for in this study (e.g., eating while standing or weight-shifting), may also require separate labelling. Other solutions may include employing more sensitive means of validating weight shifts that do not rely on visual classification (e.g., force plates),⁶³ accounting for the behavioural states over a longer time period preceding and following standing, or investigating alternatives such as using a single wither sensor to detect mediolateral postural sway as an indicator of subtle weight shifts in limb pairs as has been used to identify horses with constrained postural control.⁴² As well as investigating associations between occurrence of specific behaviours with orthopaedic pain in future studies, transition and variability of behaviour may also be important predictors of pain, warranting further investigation.

Our study showed higher sensitivity and precision than studies using 10-s intervals, except for walking which was comparable, most likely because walking can be a continuous behaviour and easily averaged over 10 s. We chose to classify behaviour at a time-sampling interval of 1 s attempting to obtain the highest sensitivity for subtle movements. Intervals of 1 s were used in dogs to achieve >95% accuracy for six of eight behaviours.³⁷ The combination of sampling rate and time interval used can influence the accuracy of detecting horse behaviour.⁵⁵ Eerdeken et al. showed an increase of time interval from 0.6 to 1.2 s resulted in significant improvements to predictive performance.⁵⁵ Robert et al. compared the accuracy of 3-, 5- and 10-s intervals to determine behaviour in cattle and found agreement between sensor and observation was higher in 3- (98.1%) and 5-s intervals (97.7%) compared with the use of a 10-s interval (85.4%).⁴⁴ High variations in readings recorded over 10-s intervals were thought to reduce accuracy.

This study had some limitations. Only one observer manually labelled the defined behaviours from video observation. Future studies should consider additional observers when labelling data to reduce errors associated with subjectivity and human perception. However, previous observational studies have assessed both intra- and inter-observer reliability with positive results; 97.4% intra-observer reliability (agreement) for one observer recording recumbency behaviours in cattle,⁵³ and no significant difference was found between observations of horses recumbency recorded by two observers.⁵⁴ Second, the accelerometer and gyroscope used in this study had a battery life of less than 5 h. Although adequate for validation purposes, this would be a limiting factor for the clinical applicability of its purpose as a 24-h monitoring system. Further, data were not collected from ~02:00 to 10:00, and avoided during active periods in the yard, thus different patterns of behaviours may be observed during these times. The algorithm was not sensitive enough to accurately identify stepping, and weight-shifting and pawing, as separate progressive and

nonprogressive movements, respectively. Previous studies showed accuracy was only reduced by 5% when sampling rate was reduced from 200 to 25 Hz.⁵⁵ A reduced sampling rate would improve battery performance, but this would require further algorithm development. Investigation of the long-term use of IMU boots may require assessment for practical use. Finally, we used six horses in the development of the algorithm, and thus increasing the number of horses and types of behaviours, particularly those behaviours that are more varied and less prevalent, may improve predictions.

IMUs have the potential to objectively quantify behaviour of stabled racehorses, and with further validation, may be generalisable to the wider equestrian industry. Although the association between each behaviour and musculoskeletal injury is still unknown, the ability to document behaviour over time may allow the identification of patterns indicative of pathology. The overall performance of the algorithm we developed to determine horse activity was reasonable, but only for prevalent behaviour. Further algorithm development is required for infrequent behaviours such as pawing, and distinguishing distinct bouts of weight-shifting from those that are due to a transition to another behaviour.

AUTHOR CONTRIBUTIONS

Katrina Anderson conducted data collection and collation. Katrina Anderson, Ashleigh V. Morrice-West and Peta L. Hitchens contributed to the data analysis. All authors contributed to the study design, interpretation of results, preparation of the manuscript and gave final approval.

ACKNOWLEDGEMENTS

The authors thank the team at Logemas for providing the IMU sensors and ongoing technical support; Susanne Ellens, Victoria University, for her assistance during the early stage of algorithm development; and the owners and trainers of the horses that participated in the study. Jointly funded by Racing Victoria, the Victorian Racing Industry Fund of the Victoria State Government and The University of Melbourne. K. Anderson was funded by a University of Melbourne Graduate Research Scholarship. Open access publishing facilitated by The University of Melbourne, as part of the Wiley - The University of Melbourne agreement via the Council of Australian University Librarians.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study, excluding video recordings that are identifiable, are available from the corresponding author upon reasonable request.

ETHICS STATEMENT

The study was approved by the University of Melbourne Animal Ethics Committee (Reference number 1814523). Written consent for this study was obtained from the horse owner or duly authorised agent for the owner.

ORCID

Katrina Anderson  <https://orcid.org/0000-0001-7249-5629>
 Ashleigh V. Morrice-West  <https://orcid.org/0000-0003-2776-4273>
 Elizabeth A. Walmsley  <https://orcid.org/0000-0002-2641-7977>
 Andrew D. Fisher  <https://orcid.org/0000-0002-2505-2160>
 R. Chris Whitton  <https://orcid.org/0000-0003-0012-4065>
 Peta L. Hitchens  <https://orcid.org/0000-0002-7528-7056>

REFERENCES

- Bailey C, Rose R, Reid S, Hodgson D. Wastage in the Australian thoroughbred racing industry: a survey of Sydney trainers. *Aust Vet J.* 1997; 75(1):64–6. <https://doi.org/10.1111/j.1751-0813.1997.tb13836.x>
- Dyson PK, Jackson BF, Pfeiffer DU, Price JS. Days lost from training by two- and three-year-old thoroughbred horses: a survey of seven UK training yards. *Equine Vet J.* 2008;40(7):650–7. <https://doi.org/10.2746/042516408X363242>
- Holmes JM, Mirams M, Mackie EJ, Whitton RC. Thoroughbred horses in race training have lower levels of subchondral bone remodelling in highly loaded regions of the distal metacarpus compared to horses resting from training. *Vet J.* 2014;202(3):443–7. <https://doi.org/10.1016/j.tvjl.2014.09.010>
- Parkin TDH, Clegg PD, French NP, Proudman CJ, Riggs CM, Singer ER, et al. Catastrophic fracture of the lateral condyle of the third metacarpus/metatarsus in UK racehorses – fracture descriptions and pre-existing pathology. *Vet J.* 2006;171(1):157–65. <https://doi.org/10.1016/j.tvjl.2004.10.009>
- Riggs CM, Whitehouse GH, Boyde A. Pathology of the distal condyles of the third metacarpal and third metatarsal bones of the horse. *Equine Vet J.* 1999;31(2):140–8. <https://doi.org/10.1111/j.2042-3306.1999.tb03807.x>
- Stover SM, Murray A. The California postmortem program: leading the way. *Vet Clin North Am Equine Pract.* 2008;24(1):21–36. <https://doi.org/10.1016/j.cveq.2007.11.009>
- Tranquille CA, Murray RC, Parkin TDH. Can we use subchondral bone thickness on high-field magnetic resonance images to identify thoroughbred racehorses at risk of catastrophic lateral condylar fracture? *Equine Vet J.* 2017;49(2):167–71. <https://doi.org/10.1111/evj.12574>
- Trope GD, Ghasem-Zadeh A, Anderson GA, Mackie EJ, Whitton RC. Can high-resolution peripheral quantitative computed tomography imaging of subchondral and cortical bone predict condylar fracture in thoroughbred racehorses? *Equine Vet J.* 2015;47(4):428–32. <https://doi.org/10.1111/evj.12312>
- Kawcak CE, McIlwraith CW, Norrdin RW, Park RD, Steyn PS. Clinical effects of exercise on subchondral bone of carpal and metacarpophalangeal joints in horses. *Am J Vet Res.* 2000;61(10):1252–8.
- Crijns CP, Martens A, Bergman HJ, Veen H, Duchateau L, Bree HJJ, et al. Intramodality and intermodality agreement in radiography and computed tomography of equine distal limb fractures. *Equine Vet J.* 2014;46(1):92–6. <https://doi.org/10.1111/evj.12082>
- Racing Australia. *Australian Racing Fact Book 2007/2008.*
- Racing Australia. *Australian Racing Fact Book 2017/2018.*
- Torcivia C, McDonnell S. In-person caretaker visits disrupt ongoing discomfort behavior in hospitalized equine orthopedic surgical patients. *Animals.* 2020;10(2):210. <https://doi.org/10.3390/ani10020210>
- Rietmann TR, Stauffacher M, Bernasconi P, Auer JA, Weishaupt MA. The association between heart rate, heart rate variability, endocrine and behavioural pain measures in horses suffering from laminitis. *J Vet Med A Physiol Pathol Clin Med.* 2004;51(5):218–25. <https://doi.org/10.1111/j.1439-0442.2004.00627.x>
- Bussi eres G, Jacques C, Lainay O, Beauchamp G, Leblond A, Cadore J-L, et al. Development of a composite orthopaedic pain

- scale in horses. *Res Vet Sci.* 2008;85(2):294–306. <https://doi.org/10.1016/j.rvsc.2007.10.011>
16. Price J, Catriona S, Welsh EM, Waran NK. Preliminary evaluation of a behaviour based system for assessment of post operative pain in horses following arthroscopic surgery. *Vet Anaesth Analg.* 2003; 30(3):124–37. <https://doi.org/10.1046/j.1467-2995.2003.00139.x>
 17. Pritchett LC, Ulibarri C, Roberts MC, Schneider RK, Sellon DC. Identification of potential physiological and behavioral indicators of postoperative pain in horses after exploratory celiotomy for colic. *Appl Anim Behav Sci.* 2003;80(1):31–43. [https://doi.org/10.1016/S0168-1591\(02\)00205-8](https://doi.org/10.1016/S0168-1591(02)00205-8)
 18. Dalla Costa E, Minero M, Lebel D, Stucke D, Canali E, Leach MC. Development of the Horse Grimace Scale (HGS) as a pain assessment tool in horses undergoing routine castration. *PLoS One.* 2014;9(3): e92281. <https://doi.org/10.1371/journal.pone.0092281>
 19. Dyson S, Berger JM, Ellis AD, Mullard J. Can the presence of musculoskeletal pain be determined from the facial expressions of ridden horses (FEReq)? *J Vet Behav Clin Appl Res.* 2017;19:78–89. <https://doi.org/10.1016/j.jveb.2017.03.005>
 20. Glerup KB, Forkman B, Lindegaard C, Andersen PH. An equine pain face. *Vet Anaesth Analg.* 2015;42(1):103–14. <https://doi.org/10.1111/vaa.12212>
 21. Hewetson M, Christley RM, Hunt ID, Voute LC. Investigations of the reliability of observational gait analysis for the assessment of lameness in horses. *Vet Rec.* 2006;158(25):852–8. <https://doi.org/10.1136/vr.158.25.852>
 22. Lindegaard C, Thomsen MH, Larsen S, Andersen PH. Analgesic efficacy of intra-articular morphine in experimentally induced radiocarpal synovitis in horses. *Vet Anaesth Analg.* 2010;37(2):171–85. <https://doi.org/10.1111/j.1467-2995.2009.00521.x>
 23. van Loon JP, Van Dierendonck MC. Monitoring acute equine visceral pain with the equine Utrecht university scale for composite pain assessment (EQUUS-COMPASS) and the equine Utrecht university scale for facial assessment of pain (EQUUS-FAP): a scale-construction study. *Vet J.* 2015;206(3):356–64. <https://doi.org/10.1016/j.tvjl.2015.08.023>
 24. Rashid M, Silventoinen A, Glerup KB, Andersen PH. Equine facial action coding system for determination of pain-related facial responses in videos of horses. *PLoS One.* 2020;15(11):e0231608. <https://doi.org/10.1371/journal.pone.0231608>
 25. van Loon J, Van Dierendonck M. Objective pain assessment in horses (2014–2018). *Vet J.* 2018;242:1–7. <https://doi.org/10.1016/j.tvjl.2018.10.001>
 26. Ask K, Rhodin M, Tamminen L-M, Hernelund E, Haubro AP. Identification of body behaviors and facial expressions associated with induced orthopedic pain in four equine pain scales. *Animals.* 2020;10(11): 2155. <https://doi.org/10.3390/ani10112155>
 27. Broomé S, Ask K, Rashid-Engström M, Haubro Andersen P, Kjellström H. Sharing pain: using pain domain transfer for video recognition of low grade orthopedic pain in horses. *PLoS One.* 2022; 17(3):e0263854. <https://doi.org/10.1371/journal.pone.0263854>
 28. Lieber B, Taylor BES, Appelboom G, McKhann G, Connolly ES. Motion sensors to assess and monitor medical and surgical management of parkinson disease. *World Neurosurg.* 2015;84(2):561–6. <https://doi.org/10.1016/j.wneu.2015.03.024>
 29. Lugade V, Fortune E, Morrow M, Kaufman K. Validity of using tri-axial accelerometers to measure human movement – part I: posture and movement detection. *Med Eng Phys.* 2014;36(2):169–76. <https://doi.org/10.1016/j.medengphy.2013.06.005>
 30. Mathie MJ, Coster ACF, Lovell NH, Celler BG. Detection of daily physical activities using a triaxial accelerometer. *Med Biol Eng Comput.* 2003;41(3):296–301. <https://doi.org/10.1007/bf02348434>
 31. Mathie MJ, Celler BG, Lovell NH, Coster ACF. Classification of basic daily movements using a triaxial accelerometer. *Med Biol Eng Comput.* 2004;42(5):679–87. <https://doi.org/10.1007/bf02347551>
 32. Aghanavasi S, Bergquist F, Nyholm D, Senek M, Memedi M. Motion sensor-based assessment of Parkinson's disease motor symptoms during leg agility tests: results from levodopa challenge. *IEEE J Biomed Health Inform.* 2020;24(1):111–9. <https://doi.org/10.1109/jbhi.2019.2898332>
 33. Elliott KH, Vaillant ML, Kato A, Speakman JR, Ropert-Coudert Y. Accelerometry predicts daily energy expenditure in a bird with high activity levels. *Biol Lett.* 2013;9(1):20120919. <https://doi.org/10.1098/rsbl.2012.0919>
 34. Fehlmann G, O'Riain MJ, Hopkins PW, O'Sullivan J, Holton MD, Shepard ELC, et al. Identification of behaviours from accelerometer data in a wild social primate. *Anim Biotelemetry.* 2017;5(1):6. <https://doi.org/10.1186/s40317-017-0121-3>
 35. Graf PM, Wilson RP, Qasem L, Hackländer K, Rosell F. The use of acceleration to code for animal behaviours; a case study in free-ranging eurasian beavers *castor fiber.* *PLoS One.* 2015;10(8):e0136751. <https://doi.org/10.1371/journal.pone.0136751>
 36. Leos-Barajas V, Photopoulou T, Langrock R, Patterson TA, Watanabe YY, Murgatroyd M, et al. Analysis of animal accelerometer data using hidden markov models. *Methods Ecol Evol.* 2017;8(2): 161–73. <https://doi.org/10.1111/2041-210X.12657>
 37. den Uijl I, Gómez Álvarez CB, Bartram D, Dror Y, Holland R, Cook A. External validation of a collar-mounted triaxial accelerometer for second-by-second monitoring of eight behavioural states in dogs. *PLoS One.* 2017;12(11):e0188481. <https://doi.org/10.1371/journal.pone.0188481>
 38. Barwick J, Lamb D, Dobos R, Schneider D, Welch M, Trotter M. Predicting lameness in sheep activity using tri-axial acceleration signals. *Animals.* 2018;8(1):12. <https://doi.org/10.3390/ani8010012>
 39. Gibbons J, Medrano-Galarza C, Marie de Passillé A, Rushen J. Lying laterality and the effect of icetag data loggers on lying behaviour of dairy cows. *Appl Anim Behav Sci.* 2012;136(2):104–7. <https://doi.org/10.1016/j.applanim.2011.12.005>
 40. Bachmann M, Wensch-Dorendorf M, Hoffmann G, Steinhöfel I, Bothendorf S, Kemper N. Pedometers as supervision tools for mares in the prepartal period. *Appl Anim Behav Sci.* 2014;151:51–60. <https://doi.org/10.1016/j.applanim.2013.11.014>
 41. Steinke SL, Montgomery JB, Barden JM. Accelerometry-based step count validation for horse movement analysis during stall confinement. *Front Vet Sci.* 2021;8(680):681213. <https://doi.org/10.3389/fvets.2021.681213>
 42. Egan S, Brama PAJ, Goulding C, McKeown D, Kearney CM, McGrath D. The feasibility of equine field-based postural sway analysis using a single inertial sensor. *Sensors.* 2021;21(4):1286. <https://doi.org/10.3390/s21041286>
 43. Trénel P, Jensen MB, Decker EL, Skjøth F. Quantifying and characterizing behavior in dairy calves using the icetag automatic recording device. *J Dairy Sci.* 2009;92(7):3397–401. <https://doi.org/10.3168/jds.2009-2040>
 44. Robert B, White BJ, Renter DG, Larson RL. Evaluation of three-dimensional accelerometers to monitor and classify behavior patterns in cattle. *Comput Electron Agric.* 2009;67(1):80–4. <https://doi.org/10.1016/j.compag.2009.03.002>
 45. White BJ, Coetzee JF, Renter DG, Babcock AH, Thomson DU, Andresen D. Evaluation of two-dimensional accelerometers to monitor behavior of beef calves after castration. *Am J Vet Res.* 2008; 69(8):1005–12. <https://doi.org/10.2460/ajvr.69.8.1005>
 46. Whiteside D, Cant O, Connolly M, Reid M. Monitoring hitting load in tennis using inertial sensors and machine learning. *Int J Sports Physiol Perform.* 2017;12(9):1212–7. <https://doi.org/10.1123/ijspp.2016-0683>
 47. Ungar ED, Nevo Y, Baram H, Arieli A. Evaluation of the icetag leg sensor and its derivative models to predict behaviour, using beef cattle on rangeland. *J Neurosci Methods.* 2018;300:127–37. <https://doi.org/10.1016/j.jneumeth.2017.06.001>

48. StataCorp. 2017. Stata Statistical Software: Release 15. College Station, TX: StataCorp LLC.
49. Martiskainen P, Järvinen M, Skön J-P, Tiirikainen J, Kolehmainen M, Mononen J. Cow behaviour pattern recognition using a three-dimensional accelerometer and support vector machines. *Appl Anim Behav Sci.* 2009;119(1):32–8. <https://doi.org/10.1016/j.applanim.2009.03.005>
50. Gabadinho A, Ritschard G, Müller NS, Studer M. Analyzing and visualizing state sequences in R with TraMineR. *J Stat Softw.* 2011;40(4): 1–37. <https://doi.org/10.18637/jss.v040.i04>
51. R Core Team (2017). R: A language and environment for statistical computing. Vienna: R Foundation for Statistical Computing. <http://www.R-project.org/>
52. Chung ELT, Khairuddin NH, Azizan TRPT, Adamu L. Sleeping patterns of horses in selected local horse stables in Malaysia. *J Vet Behav.* 2018;26:1–4. <https://doi.org/10.1016/j.jveb.2018.03.014>
53. Ledgerwood DN, Winckler C, Tucker CB. Evaluation of data loggers, sampling intervals, and editing techniques for measuring the lying behavior of dairy cattle. *J Dairy Sci.* 2010;93(11):5129–39. <https://doi.org/10.3168/jds.2009-2945>
54. DuBois C, Zakrajsek E, Haley DB, Merkies K. Validation of triaxial accelerometers to measure the lying behaviour of adult domestic horses. *Animal.* 2015;9(1):110–4. <https://doi.org/10.1017/s175173111400247x>
55. Eerdeken A, Deruyck M, Fontaine J, Martens L, Poorter ED, Joseph W. Automatic equine activity detection by convolutional neural networks using accelerometer data. *Comput Electron Agric.* 2020; 168:168105139. <https://doi.org/10.1016/j.compag.2019.105139>
56. Thompson CJ, Luck LM, Keshwani J, Pitla SK, Karr LK. Location on the body of a wearable accelerometer affects accuracy of data for identifying equine gaits. *J Equine Vet.* 2018;63:1–7. <https://doi.org/10.1016/j.jjevs.2017.12.002>
57. González LA, Bishop-Hurley GJ, Handcock RN, Crossman C. Behavioral classification of data from collars containing motion sensors in grazing cattle. *Comput Electron Agric.* 2015;110:91–102. <https://doi.org/10.1016/j.compag.2014.10.018>
58. Hämäläinen WM, Martiskainen P, Järvinen M, Skön J-P, Tiirikainen J, Kolehmainen M, et al. Computational challenges in deriving dairy cows' action patterns from accelerometer data. *Siilinjärvi, Finland: Finnish Society for Applied Ethology;* 2010. p. 18. <https://orgprints.org/id/eprint/17743/1/isaeb2010.pdf>
59. Smith D, Rahman A, Bishop-Hurley GJ, Hills J, Shahriar S, Henry D, et al. Behavior classification of cows fitted with motion collars: decomposing multi-class classification into a set of binary problems. *Comput Electron Agric.* 2016;131:40–50. <https://doi.org/10.1016/j.compag.2016.10.006>
60. Sweet AL. Temporal discrimination by the human eye. *Am J Psychol.* 1953;66:185–98. <https://doi.org/10.2307/1418725>
61. Ishihara A, Bertone AL, Rajala-Schultz PJ. Association between subjective lameness grade and kinetic gait parameters in horses with experimentally induced forelimb lameness. *Am J Vet Res.* 2005; 66(10):1805–15. <https://doi.org/10.2460/ajvr.2005.66.1805>
62. McCracken MJ, Kramer J, Keegan KG, Lopes M, Wilson DA, Reed SK, et al. Comparison of an inertial sensor system of lameness quantification with subjective lameness evaluation. *Equine Vet J.* 2012;44(6): 652–6. <https://doi.org/10.1111/j.2042-3306.2012.00571.x>
63. Clayton HM, Nauwelaerts S. Is a single force plate adequate for stabilographic analysis in horses? *Equine Vet J.* 2012;44(5):550–3. <https://doi.org/10.1111/j.2042-3306.2011.00458.x>

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Anderson K, Morrice-West AV, Walmsley EA, Fisher AD, Whitton RC, Hitchens PL. Validation of inertial measurement units to detect and predict horse behaviour while stabled. *Equine Vet J.* 2023. <https://doi.org/10.1111/evj.13909>